MIGUEL ÂNGELO PARDAL JOÃO CARLOS MARQUES MANUEL AUGUSTO GRAÇA Scientific Editors

Aquatic Ecology of the Mondego River Basin Global Importance of Local Experience





Coimbra • Imprensa da Universidade

MIGUEL ÂNGELO PARDAL JOÃO CARLOS MARQUES MANUEL AUGUSTO GRAÇA Scientific Editors

Aquatic Ecology of the Mondego River Basin Global Importance of Local Experience



Coimbra • Imprensa da Universidade

COORDENAÇÃO EDITORIAL Imprensa da Universidade de Coimbra

> CONCEPÇÃO GRÁFICA António Barros

INFOGRAFIA António Resende Estímulus [design] • Coimbra

> Execução gráfica GRAFIASA

ILUSTRAÇÃO DA CAPA P. P. Cunha e J. Dinis

> ISBN 972-8704-04-6

DEPÓSITO LEGAL 175038/02

© JANEIRO 2002, IMPRENSA DA UNIVERSIDADE DE COIMBRA





OBRA PUBLICADA COM O PATROCÍNIO DE: IMAR – INSTITUTO DO MAR IPIMAR – INSTITUTO DE INVESTIGAÇÃO DAS PESCAS E DO MAR

NITROGEN DYNAMICS IN THE MONDEGO ESTUARY: LEAF SENESCENCE AND N MOBILISATION IN Spartina maritima

Abstract

Spartina maritima (Curtis) Fernald is a dominant species in the Mondego salt marsh playing a significant role in estuarine productivity and energy cycling. A study conducted between February 1996 and January 1997 revealed that nitrate was mainly transported from roots and rhizomes to the leaves where it was accumulated. Nitrate concentration increased during summer in all types of studied leaves, probably playing an important role in osmotic regulation. Ammonium, on the contrary, was incorporated into organic compounds in the root rather than transported into the leaves. Total nitrogen quantified in the leaves, showed an obvious pattern, decreasing greatly during summer and early autumn. From March to June, nitrogen content in green leaves decreased nearly 75 %. This variation of nitrogen content was highly correlated with its concentration in the sediment. Leaf ageing in *Spartina maritima* was associated to chlorophyll losses, to nitrate and total nitrogen remobilization, and also to ammonium accumulation.

Introduction

In salt marshes, nitrogen supply is mediated by redox of the sediments and limits plant growth (Valiela 1995). Nitrate and ammonium are the major sources of inorganic nitrogen taken up by roots of *Spartina alterniflora* as well as in general higher plants (Wirén et al. 1997). Nitrate is readily mobilised in xylem and can be stored in vacuoles of roots, shoots, and storage organs (Martinoia et al. 1981). Its accumulation in vacuoles is of considerable importance for cation-anion balance and osmoregulation (Smirnoff and Stewart 1985). Ammonium is incorporated into organic compounds and generally not stored in cells (Schortemeyer et al. 1997).

313

Spartina maritima is a dominant species in the Mondego salt marsh (Portugal) and plays an important role in nutrient and energy dynamics in European salt marshes similar to that played by Sparting alterniflorg in North American marshes. Like most

⁽¹⁾ IMAR – Instituto do Mar, Centro Interdisciplinar de Coimbra, Departamento de Botanica, Universidade de Coimbra, 3000 Coimbra, Portugal

coastal producers, throughout the world Spartina maritima is subject to eutrophication, which elevates nitrogen levels in the sediment (Valiela et al. 1992). Increased nitrogen supply increases growth and biomass, and affects plant morphology, nutrient content, density and reproduction (Rogers et al. 1998).

In general, plant leaves undergo a series of developmental changes. Initially, leaves are predominantly heterotrophic organs, maturing to become a net exporter of photosynthates. After reaching maturity, the leaf exhibits a reduced capacity to act as a source of photosynthetically fixed carbon and enters a senescent phase (Turgeon 1989). Leaf senescence represents a key developmental phase in plant life, which is as ordered and complex as any other phase of development. During leaf senescence, N, C and minerals are mobilised from the mature leaf to other parts of the plant. This process involves a series of events like cessation of photosynthesis, disintegration of chloroplasts, breakdown of proteins, loss of chlorophyll and removal of amino acids (Buchanan-Wollaston 1997).

The main objective of this work was to estimate seagrass contribution to the nitrogen dynamics in a Portuguese saltmarsh. This paper reports the seasonal variation of inorganic and total nitrogen concentrations in the Mondego saltmarsh and *Spartina maritima* contribution to this process. Nitrogen mobilisation during leaf senescence and its importance for plant survival in this system was also studied.

Material and methods

Study site

314



This work was conducted at the Mondego estuary located on the western coast of Portugal (40°08' N / 8°50' W) (Fig. 1). The Mondego estuary consists of two arms

I. The Mondego estuary.

with different hydrographic characteristics. The northern arm is deeper, while the southern arm is largely silted up, routing most of the freshwater input through the northern arm. Water circulation in the southern arm is mainly dependent on tidal cycles, receiving also small freshwater inputs of the Pranto river, a tributary with large amounts of nitrogen substances, frequently used as fertilisers by rice farmers in upstream areas (Marques 1989, Flindt et al. 1997). Sampling site was located on the south arm, in a salt marsh dominated by *Sparting maritima* (Fig. 1A).

Field program and laboratory procedures

Spartina maritima plants were sampled monthly from February 1996 to January 1997. In the laboratory, biomass samples were separated into roots, rhizomes, and leaves. Leaves were separated in 3 leaf age groups: green leaves (GL) – live tissue with green colour; senescent leaves (SL1 and SL2) – senescent tissue with at least 1/3 of area yellow (SL1), leaves with almost 2/3 of their area with yellow-brown color (SL2) and standing-decaying leaves (SDL) – yellow-brown dead material ready to fall. Leaves detached via decay were collected using 20 nylon mesh litter traps 100 cm long x 40 cm wide, opened at the top for monthly collections (Castro and Freitas 2000), designated in this paper as litter trap leaves (LTL). Samples were weighed and dried at 60°C until constant weight. Dry tissues were ground and analysed for NO₃ by the acid salicylic method (Cataldo et al. 1975), for NH4* by the indophenol blue method (Solorzano 1969), and for total N (only in leaf samples) by the Kjeldhal method (Bremner and Mulvney 1982).

Immediately after collection, the chlorophyll (a + b) content was measured spectrophotometrically on triplicate 300 mg (fresh weight) of leaf samples (GL, SL1, SL2 and SDL), after extraction with 80 % acetone (Henry and Price 1993).

Sediment samples were collected each month in parallel to plant samples. Sediments were oven-dried at 60°C for 3 days and stored in sterilised plastic bags. Total N was determined in 5 g of sediment following the method in Bonneau and Souchier (1979).

Statistical analysis

One Way Analysis o Variance (ANOVA) was used to test for differences between mean nitrogen concentrations in plant samples, chlorophyll content in leaves, and seasonal variation of these variables. Non-parametric analysis of variance (Kruskal-Wallis) was used whenever parametric assumptions were not verified (Zar 1996).

315

Results

Water content

Water content (percentage of fresh weight) was higher in belowground tissues than in leaves (Fig. 2). Water content was relatively constant in roots and rhizomes, with higher values in root tissues. Rhizomes showed a medium annual value of 76 % of water and roots of 84 %. Young leaves were more succulent than senescent and dead leaves. All types of leaves revealed a decrease in water content from February to April, increasing the month after. Green and senescent leaves showed a slight decrease during summer. Water content in standing dead leaves increased from May to October, remaining high in winter.



Fig. 2. Water content, expressed as percentage of fresh weight, in Spartina mantima leaves, roots and rhizomes, collected from February 1996 to January 1997.

316

Chlorophyll

Chlorophyll levels were different between the four types of leaves ($H_{3,N=144}=123$; P<0.001; Kruskal-Wallis ANOVA), decreasing with leaf senescence (Fig. 3). Overall, mean chlorophyll concentration for GL was 24.5 ± 3.1 mg L⁴. Mean losses of 17 %, 39 % and 58 % were measured for SL1, SL2 and SDL, respectively. Although each type of leaf followed its own seasonal change, a late-summer decrease and autumn increase were common features of the annual pattern.



Fig. 3. Chlorophyll content (a + b) in leaves of Spartina maritima, monthly collected from February 1996 to January 1997.

Nitrogen dynamics

Nitrate was clearly transported and accumulated in young leaves (Fig. 4). During the study period, belowground roots and rhizomes showed levels of NO₃⁻ 4-5 times lower than GL Seasonal variations were observed in all types of leaves, with higher



Fig. 4. Nitrate concentrations determined in plant samples between February 1996 and January 1997.

values in summer months. Differences between root and rhizome NO₃⁻ concentrations were verified (F₁₂₁₉=7.46; P<0,01). Seasonal variations for belowground tissues were also observed with slight increase in dry months. NO₃⁻ mean concentrations in rhizomes ranged from 6.4 \pm 1.7 µmol g⁻¹ dry wt (February) to 25.0 \pm 12.8 µmol g⁻¹ dry wt (June). Nitrate leaf content changed greatly with leaf age (F₄₃₇₄=49.78; P<0,001). Higher concentrations consistently occurred in the youngest age class, with large decrement from mature to senescent leaves. Leaves detached via decay (LTL) revealed about 70 % less nitrate than GL.

Mean monthly concentrations of NH4⁺ increased with leaf senescence (F_{4,175} = 14.35; P<0,05). Like NO3⁻, all types of leaves showed NH4⁺ seasonal changes (Fig. 5). A slight decrease in NH4⁺ content was observed in leaves collected during summer. Standing-decaying leaves (SDL) showed ammonium concentrations 2-3 times higher than GL. Similar concentrations were found in belowground tissues, and roots and rhizomes did not show statistical differences in their values (F_{1,70} = 0.52; P=0.47).



Fig. 5. Ammonium concentrations determined in plant samples between February 1996 and January 1997.

All leaves revealed a strong seasonal variation, with leaves produced in early spring and winter showing higher N levels (Fig. 6). Total nitrogen content (TN) was significantly different between the types of leaves analysed (F4155 = 6.94; P<0.05). Mean N concentrations in GL and SL1 were generally higher from that observed in older leaves from February to June. This difference decreased and was even inverted during summer and autumn, recovering in the last month.



Fig. 6. Total nitrogen content measured in leaves of Spartina maritima between February 1996 and January 1997.

Total nitrogen (TN) concentrations in the sediment were much higher in spring and winter (Fig. 7). Maximum value of 20 ± 0.5 mmol g⁻¹ dry wt was measured in April. In June, total nitrogen concentrations decreased to 5 ± 1.1 mmol g⁻¹ dry wt.



Fig. 7. Seasonal changes in nitrogen concentrations in sediment samples, collected in the Mondego salt marsh from February 1996 to January 1997.

Discussion

Nitrate translocation from roots to leaves (Fig. 4) can be explained by the need of tissue osmotic adjustment, particularly important during summer, when soil salinity increases (Flowers et al. 1986). The accumulation of salts participate in this function and is the least energetic costly mechanism of osmolarity generation (Raven 1985). Water content in *Spartina maritima* leaves did not change greatly along the year in living leaves, suggesting that succulence is not an important strategy for tissue osmoregulation (Fig. 2). Considering that, contrarily to dicotiledone plants (Flowers et al. 1986), grasses do not use succulence as a major adaptation to soil salinity, nitrate can thus be considered to play an important role in this process for this species. Our results, particularly the NO₃⁻ accumulation in summer leaves, showed that NO₃⁻ can also work as an important mechanism to lower the energy costs of the plant for nitrate reduction and assimilation in these tissues, since the energy can be directly given by the photosynthesis process (Abrol et al. 1983), and also decreasing the production of toxic oxygen radicals. Several studies demonstrate that light increases nitrate metabolism rates (Delhon et al. 1995, Keller et al. 1995, Cárdenas-Navarro et al. 1998).

Toxic ammonium effects are well known in terrestrial plants (Gerendás et al. 1997, Pearson and Stewart 1993), but for marine environments we can expect to have higher NH⁴⁺ concentrations in plant tissues, since salt concentration in sediments is much higher, and plants are probably more tolerant to this nutrient. However, this was not observed. Whereas NO₂⁻⁻ was stored in leaves without detrimental effect, NH⁴⁺ content was quite low in both aerial and subterranean organs. This showed that, despite the high NH⁴⁺ concentrations in the soil (Fig. 5), Spartina maritima has several physiological mechanisms to prevent the increase of this nutrient in its tissues. Van Katwijk et al (1997) showed that the presence of concentrations as low as 25 μ M of NH⁴⁺ in sediments were toxic to Zostera marina plants. Fan et al. (1997) also suggested that the use of NO₃⁻⁻ instead of NH⁴⁺ as an N source is important for rice plants to tolerate anaerobiosis conditions of the soil, particularly for seed germination, because it gives a more efficient economy of energy and minimises lower pH effects in cells cytoplasm.

The lower N concentrations observed in summer were expected because the flowering period of this species occurs between May and July, and nutrients become thus necessary for plant reproduction (Marschner 1995). What is interesting is that the N seasonal variation in the soil corresponds almost exactly to the same variation in the sediment (Fig. 7) reflecting an important strategy that *Sparting maritimg* plants use to improve nutritional efficiency during low external N availability (Thornton et al. 1994). During summer a decline of N absorption at root level is expected (Correia et al. 1992), mainly due to the reduction of carbohydrate levels in root cells (Marschner 1995).

Ammonium concentrations have been shown to increase during leaf senescence for several species (e.g. Thomas 1978, Peters and Van Laere 1992, Chen et al. 1997). This study showed that it also happens in *Spartina maritima* (Fig. 5). A decrease in glutamine synthetase activity, breakdown of macromolecules (e.g., proteins and nucleic acids) and an increase in nitrate reduction during leaf ageing, are some of the factors controlling ammonium accumulation (Storey and Beevers 1978, Platt and Anthon

1981, Kar and Feierabend 1984, Chen and Kao 1998). Chen et al. (1997) suggested that NH₄⁺ could participate in senescence regulation of rice leaves, with the argument that increases in ammonium concentrations could make foliar tissues more sensitive to the action of ethylene, thus promoting senescence.

An opposite pattern was observed for nitrate and total N; senescent leaves clearly showed lower nitrate concentrations than green leaves. Castro and Freitas (2000) showed that higher leaf fall rates of *Spartina maritima* occur during summer, peaking in July and that these results are clearly related to the decrease of N and NOs⁻ in leaves before they fall, contributing to nutrient and energy saving of the plant (Marschner 1995, Hayati et al. 1995).

Conclusions

Leaf senescence and decomposition are important processes for the cycling of nutrients in the salt marsh system (Cornelissen 1996, Rutigliano et al. 1996, Aerts and Caluwe 1997, Mugendi and Nair 1997). Nutrient regulation, particularly nitrogen, is a very important process during plant development (Marschner et al. 1997). This work showed that *Spartina maritima* was capable of adjusting N distribution when growing in low N conditions and also before leaf fall in order to avoid the waste of N and energy, indispensable for plant survival in salt marsh systems. Nitrate and total N content decreased greatly with leaf senescing and before major leaf fall occurred. NH₄⁺ was not accumulated in plant cells, and it's concentration increased with leaf ageing, resulting from the normal process of senescence, but most likely to have an important role in controlling such process.

Acknowledgements

The Portuguese Foundation for Science and Technology supported this study (grant: PRAXIS XXI/BM/8091/96). We thank colleagues of IMAR for laboratory assistance, and soil analysis. We also thank Prof. Ivan Valiela (Boston University) for is comments on the manuscript. Thanks are extended to the anonymous reviewer for his comments and suggestions.

References

Abrol, Y. P., Sawhney, S. K. and Naik, M. S. 1983. Light and dark assimilation of nitrate in plants. Plant. Cell Environ. 6: 535-599.

Aerts, R. and Caluwe, H. 1997. Nutritional and plant-mediated controls on leaf litter decomposition of Carex species. Ecology 78: 244-260.

Bonneau, M. and Souchier, B. 1979. Pédologie. 2. Constituants et proprietés du soil. Masson Paris.

Bremner, J. M. and Mulvaney, C. S. 1982. Nitrogen-Total. In Methods of soil analysis-chemical and microbiological properties. Agronomy 9 (Part 2). 2nd ed. pp. 595-624.

Buchanan-Wollaston, V. 1997. The molecular biology of leaf senescence. J. Exp. Bot. 48: 181-199.

Cárdenas-Navarro, R., Adamowicz, S and Robin, P. 1998. Diurnal nitrate uptake in young tomato (Lycopersicon esculentum Mill.) plants: test of a feedback-based model. J. Exp. Bot. 49 (321): 721-730.

- Castro, P. and Freitas, H. 2000. Fungal biomass, and decomposition in Spartina maritima leaves in the Mondego salt marsh (Portugal). Hydrobiologia 428:171-177.
- Cataldo, D. A, Haroon, M., Schrader, L. E. and Youngs, V. L. 1975. Rapid colorimetric determination of nitrate in plant tissue by nitration of salicylic acid. Commun. Soil Sci. Plant Anal. 6: 71–80.
- Chen, S. J. and Kao, C. H. 1998. Methyl jasmonate, ammonium and leaf senescence in rice. J. Plant Physiol. 152: 353-357.
- Chen, S. J., Hung, K.T. and Kao, C. H. 1997. Ammonium accumulation is associated with senescence of rice leaves. Plant Growth Regulation 21: 195-201.
- Cornelissen, J. H. C. 1996. An experimental comparison of leaf decomposition rates in a wide range of temperate plant species and types. J. Ecol. 84: 573-582.
- Correia, O.A., Martins, A.C. and Catarino, F.M. 1992. Comparative phenology and seasonal foliar nitrogen variation in mediterranean species of Portugal. Ecologia Mediterranea 8: 7-18.
- Delhon, P., Gojon, A., Tillard, P and Passama, L. 1995. Diurnal regulation of NO3-- uptake in soybean plants. I. Changes in NO3-- influx, and N utilization in the plant during the day/night cycle. J. Exp. Bot. 46: 1585-1594.
- Fan, T.W-M, Higashi, R. M., Frenkiel, T.A. and Lane, A. N. 1997. Anaerobic nitrate and ammonium metabolism in flood-tolerant rice coleoptiles. J. Exp. Bot. 48: 1655-1666.
- Flindt, M. R., Kamp-Nielsen, L., Marques, J. C., Pardal, M. A., Bocci, M., Bendoricchio, G., Salomonsen, J., Nielsen, S. N. and Jorgensen, S.E. 1997. Description of the three shallow estuaries: Mondego River (Portugal), Roskilde Fjord (Denmark) and the Lagoon of Venice (Italy). Ecol. Model. 102: 17-31.
- Flowers, T. J., Hajibagheri, M. A. and Clipson, N. J. W. 1986. Halophytes. The Quart. Rev. Biol. 61: 313-337.
- Gerendás, J., Zhu, Z., Bendixen, R., Ratcliffe, R.G. and Sattelmacher, B. 1997. Physiological and biochemical processes related to ammonium toxicity in higher plants. Z. Pflanzenemahr. Bodenk. 160: 239-251.
- Hayati, R., Egli, D. B. and Crafts-Brandner, S. J. 1995. Carbon and nitrogen supply during seed filling and leaf senescence in soybean. Crop Science 35: 1063-1069.
- Henry, G. A. F. and Price, A. H. 1993. Stress indicators: chlorophylls and carotenoids. In Methods in comparative ecology. Edited by G. A. F. Henry and A. H. Price. Chapman and Hall. pp. 148-152.
- Kar, M. and Feierabend, J. 1984. Changes in the activities of enzymes involved in amino acid metabolism during senescence of detached wheat leaves. Physiol. Plant. 62: 39-44.
- Keller, M., Hess, B., Schwager, H., Scharer, H. and Koblet, W. 1995. Carbon and nitrogen partitioning in Vitis vinifera L: responses to nitrogen supply and limiting irradiance. Vitis 1: 19-26.
- Marques, J. C. 1989. Amphipoda (Crustacea) bentónicos da costa portuguesa: estudo taxonómico, ecológico e biogeográfico. Ph.D. thesis University of Coimbra. Coimbra.

Marschner, H. 1995. Mineral nutrition of higher plants. 2nd ed. Acad. Press, London.

- Marschner, H., Kirkby, E.A. and Engels, C. 1997. Importance of cycling and recycling of mineral nutrients within plants for growth and development. Botanica Acta 110: 265-273.
- Martinoia, E., Heck, V. and Wiemken, A. 1981. Vacuoles as storage compartments for nitrate in barley leaves. Nature 289: 292-294.
- Mugendi, D. N. and Nair, K. R. 1997. Predicting the decomposition patterns of tree biomass in tropical highland microregions of Kenya. Agroforestry Systems 35: 187-201.
- Noodén, L. D., Guiamét, J. J. and John, I. 1997. Senescence mechanisms. Physiol. Plant. 101: 746-753.
- Pearson, J. and Stewart, G. R. 1993. Transley review no. 56. The deposition of atmospheric ammonia and its effects on plants. New Phytol. 125: 283-305.
- Peeters, K. M. U. and Van Laere, A. J. 1992. Ammonium and amino acid metabolism in excised leaves of wheat (Triticum aestivum) senescing in the dark. Physiol. Plant. 84: 243-249
- Platt, S. G. and Anthon, G. 1981, Ammonia accumulation and inhibition of photosynthesis in methionine sulfoximine treated spinach. Plant Physiol. 67: 509-513
- Raven, J. A. 1985. Regulation of pH and generation of osmolarity in vascular plants: a cost-benefit analysis in relation to efficiency of use of energy, nitrogen and water. New Phytol. 101: 25-77.
- Rogers, J., Harris, J. and Valiela, I. 1998. Interaction of nitrogen supply, sea level rise, and elevation on species form and composition of salt marsh plants. Biol. Bull. 195: 235-237.
- Rutigliano, F. A., Santo; A. V., Berg, B., Alfani, A. and Fioretto, A. 1996. Lignin decomposition in decaying leaves of Fagus sylvatica L and needles of Abies alba Mill. Soil Biol. Biochem. 28: 101-106.

Schortemeyer, M., Stamp, P. and Feil, B. 1997. Ammonium tolerance and carbohydrate status in maize cultivars. Annals of Botany 79: 25-30.

Smirnoff, N. and Stewart, G.R. 1985. Nitrate assimilation and translocation by higher plants: comparative physiology and ecological consequences. Physiol. Plant. 64: 133-140.

Solorzano, L. 1969. Determination of ammonium in natural waters by the phenolhypoclorite method, Limnol, Oceanogr. 14: 799-801.

Storey, R. and Beevers, L. 1978. Enzymology of glutamine metabolism related to senescence and seed development in the pea (Pisum sativum L), Plant Physiol, 61: 494-500.

Thomas, H. 1978. Enzymes of nitrogen mobilization in detached leaves of Lolium temulentum during senescence. Planta 142: 161-169.

Thornton, By B., Millard, P. and Duff, E. I. 1994. Effects of nitrogen supply on the source of nitrogen used for regrowth of laminae after defoliation of four grass species. New Phytol. 128: 615-620.

Turgeon, R. 1989. The sink-source transition in leaves. Annu. Rev. Plant Physiol. Plant Mol. Biol. 40: 119-138.

Valiela, I. 1995. Marine ecological processes. 2nd ed. Springer-Verlag Inc., New York.

Valiela, I., Foreman, K., LaMontagne, M., Hersh, D., Peckol, P., DeMeo-Anderson, B., D'Avanzo, C., Babione, M., Sham, C. H., Brawley, J. and Latjha, K. 1992. Couplings of watersheds and coastal waters: sources and consequences of nutrient enrichment in Waquoit Bay, Massachusetts. Estuaries 15: 443-457.

Van Katwijk, M. M., Vergger, L. H.T., Schmitz, G. H.W. and Roelofs, J. G. M. 1997. Ammonium toxicity in eelgrass Zostera marina. Mar. Ecol. Prog. Ser. 157: 159-173.

Wirén, N., von, Gazzarrini, S. and Frommer, W. B. 1997. Regulation of nitrogen uptake in plants. Plant and Soil 196:191-199.

Zar, J. 1996. Biostatistical analysis. 3rd ed. Prentice Hall International, London.

Série

Investigação

•

Coimbra Imprensa da Universidade